ssPNA templated assembly of oligo(*p*-phenylenevinylene)s[†]

Pim G. A. Janssen,^a Nico Meeuwenoord,^b Gijs van der Marel,^b Sara Jabbari-Farouji,^c Paul van der Schoot,^c Mathieu Surin,^d Željko Tomović,^a E. W. Meijer^a and Albertus P. H. J. Schenning^{*a}

Received (in Cambridge, UK) 6th July 2009, Accepted 13th October 2009 First published as an Advance Article on the web 11th November 2009 DOI: 10.1039/b913307k

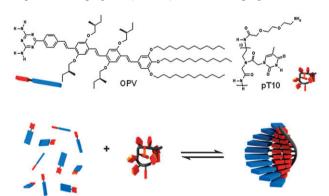
A single-stranded oligothymine peptide nucleic acid (PNA) was used as a template for the assembly of a chiral oligo-(*p*-phenylenevinylene) diaminotriazine derivative (OPV) in methylcyclohexane (MCH) revealing nanostructures in which the size is controlled by the template.

In nature, templates with specific binding sites are used to efficiently form assemblies and polymers with definite size or sequence.¹ This behavior has inspired many researchers to exploit templated polymerization^{2,3} as a tool to control the size and sequence of synthetic polymers *via* a 'bottom-up' approach.^{4–8} Especially oligonucleotides are interesting building-blocks, since they can be obtained monodisperse, functionalized and used to create predefined nanosized structures *via* sticky-end cohesion.⁹

In a previous study, we showed that the single-stranded desoxyribonucleic acid (ssDNA) oligothymine can act as a template for the assembly of complementary diaminotriazine equipped guest molecules in water.⁷ In this construct, the single DNA strand templates a supramolecular strand of chromophores held together by $\pi - \pi$, hydrophobic and hydrogen bond interactions. The efficiency of this templated assembly depends on the host-guest and guest-guest interaction and can be described by a templated assembly model based on a one-dimensional Ising model.^{7a} The use of DNA as template requires water as solvent¹⁰ and therefore the variety of guest molecules is limited. In order to broaden the scope of this templated approach to organic solvents, we now report on the use of a single-stranded peptide nucleic acid (ssPNA), consisting of 10 thymine residues (pT10, ‡Scheme 1), as a template for the assembly of a chiral π -conjugated oligo-(*p*-phenylenevinylene) diaminotriazine derivative¹¹ (**OPV**, Scheme 1) in MCH. PNA¹² is an achiral and uncharged analogue of DNA in which the phosphate backbone is replaced by an N-(2-aminoethyl)glycine backbone, making it soluble in a range of organic solvents. We have previously

† Electronic supplementary information (ESI) available: Molecular modeling. See DOI: 10.1039/b913307k shown that **OPV** forms hydrogen bonded hexamers that subsequently self-assemble into helical fibers in heptane.^{11*a*} Here, we describe the non-templated self-assembly and **pT10** templated assembly process of **OPV** studied by means of temperature-dependent UV-vis absorption and CD spectroscopy. The assemblies were visualized with atomic force microscopy (AFM).

The synthesis of OPV^{11a} and $pT10^{13}$ were performed according to literature procedures. We have first investigated the non-templated self-assembly of OPV. In chloroform, OPV is molecularly dissolved and has an absorption maxima λ_{max} at 430 nm. In MCH¹⁴ at 323 K, **OPV** (100 μ M) is molecularly dissolved since a similar absorption maximum is found. Upon cooling to 263 K, hypochromicity, a red shift of the onset of the absorption, and an absorption maximum shifting from $\lambda_{\text{max}} = 430$ to 440 nm (Fig. 1a) are observed.¹⁵ Simultaneously, at low temperatures, a positive Cotton effect is observed with a zero-crossing at $\lambda_{z-c} = 434$ nm,¹⁶ indicating that **OPV** self-assembles into right-handed helical aggregates, similar as earlier observed in heptane.¹¹ The non-templated self-assembly process has been studied in more detail by monitoring the UV absorption at $\lambda = 500$ nm as a function of temperature at different concentrations. The self-assembly is fully reversible and the observed exponential transition is indicative of a cooperative non-templated self-assembly process.^{11,17} By fitting both the concentration- and the temperature-dependent self-assembly data to the cooperative self-assembly model,¹⁷ the enthalpy of binding ($\Delta H_e \approx -75 \pm 8$ kJ mol⁻¹) was determined (Fig. 1e and f).¹⁷ To visualize the OPV assemblies, a MCH-solution has been drop-cast onto graphite (HOPG). AFM micrographs show the



Scheme 1 Molecular structures of the host template pT10 and the guest **OPV** and a schematic representation of ssPNA templated self-assembly (in blue and red OPV, and in black and red the PNA template).

^a Laboratory for Macromolecular and Organic Chemistry, Eindhoven University of Technology, P. O. Box 513, 5600 MB Eindhoven, The Netherlands. E-mail: a.p.h.j.schemning@tue.nl;

Fax: +31 40 245 1036; Tel: +31 40 247 2655

^b Laboratory of Bio-organic Synthesis, Leiden Institute of Chemistry, Gorlaeus Laboratories, Leiden University, 2300 RA Leiden, The Netherlands

^c Group Theory of Polymers and Soft Matter, P. O. Box 513, 5600 MB Eindhoven, The Netherlands

^d Laboratory for Chemistry of Novel Materials,

University of Mons-Hainaut, B-7000, Mons, Belgium

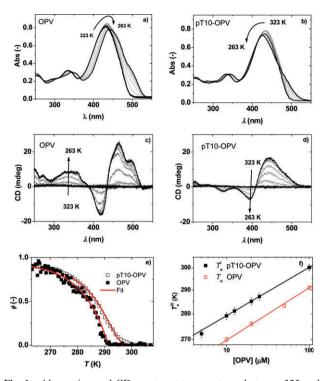


Fig. 1 Absorption and CD spectra at temperatures between 323 and 263 K for **OPV** (a and c, respectively) and **pT10–OPV** (1 : 10) (b and d, respectively) mixtures in MCH. (e) The self-assembled fraction upon cooling of **OPV** and **OPV–pT10** (10 : 1) mixtures and the fits to the cooperative self-assembly model¹⁷ and templated assembly model,^{7a} respectively. [**pT10**] = 10 μ M, [**OPV**] = 100 μ M. (f) T_e and T_e^{t} (inverted scale) as a function of [**OPV**] (logarithmic scale) for **OPV** and **pT10–OPV** (1 : 10) mixtures.

formation of fibers with a 4–6 nm height (Fig. 2a, c). This height corresponds well with the diameter of the fibers consisting of hexameric H bonded rosettes earlier reported for **OPV** in heptane.^{11*a*}

To investigate the PNA templated assembly of **OPV**, a baseequivalent of **pT10** was added to a 200 μ M solution of **OPV** in chloroform at 263 K. No Cotton effect and spectral changes were observed in the **OPV** absorption region indicating that there is no interaction between **OPV** and **pT10**. To increase the

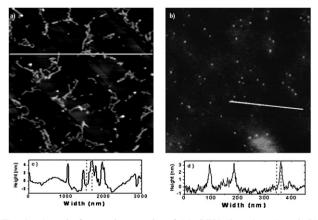


Fig. 2 Atomic force micrographs of (a) **OPV** ($3 \times 3 \mu m$) and (b) **pT10–OPV** ($1 \times 1 \mu m$) solutions drop-casted on HOPG at 273 K and the corresponding height cross-sections below. [**pT10**] = 10 μ M, [**OPV**] = 100 μ M.

host–guest and guest–guest interaction in the templated PNA assembly, MCH was used as a solvent. In this solvent **OPV** itself already forms self-assembled fibers (*vide supra*) which have to be less stable than the proposed **OPV–pT10** constructs. When **OPV** is mixed with a base-equivalent of **pT10** in MCH at 323 K and cooled down to 263 K, hypochromicity is accompanied by a blue shift of λ_{max} to 425 nm and a red shift of the onset (Fig. 1b). Compared to the non-templated **OPV** self-assembly, the **OPV–pT10** mixture has a lower intensity of the Cotton effect (Fig. 1d). Furthermore, the zero-crossing of the Cotton effect λ_{z-c} is 410 nm¹⁶ for the **pT10–OPV** mixture, while for the **OPV**, λ_{z-c} is 434 nm (Fig. 1b, c). This indicates that **OPV** is differently organized when the template **pT10** is present.¹⁸

The templated assembly process has been studied in more detail by monitoring the UV-vis absorption at $\lambda = 500$ nm as a function of temperature at different concentrations and compared to the non-templated self-assembly. The transition temperatures, below which the two types of self-assembly set in, are defined as the *elongation temperature* T_e^{17} for non-templated self-assembly and as the apparent elongation temperature T_e^{t} for templated assembly.^{7a} For a similar concentration, the T_e^t of the **pT10–OPV** mixtures is higher than the T_e of **OPV** (Fig. 1e and f), showing that the pT10-OPV assemblies are more stable than the non-templated self-assemblies of OPV. When fitting the temperaturedependent data to the templated self-assembly model as described previously,^{7a} an enthalphy of $\Delta H_e^{t} \approx -90 \pm$ 10 kJ mol⁻¹, a guest–guest interaction energy of $\varepsilon = -6.2 \pm$ $0.5 kT_{\rm p}$ was obtained.¹⁹

The enthalpy values extracted for the templated and non-templated assembly processes suggest that the higher stability of the templated assembly as indicated by the higher melting temperature is due to its larger enthalpy gain. A necessary condition for the predominance of templated assembly over self-assembly is that the free-energy change resulting from the combined effects of host-guest and templated guest-guest interaction molecules is larger than the free-energy gain from the stacking of guest molecules in self-assembly. As a consequence, the presence of the PNA template effectively suppresses the self-assembly of **OPV** unless a large excess of **OPV** is present in the solution and only then when the **pT10** templates are filled.

To visualize the **pT10–OPV** assemblies, an MCH-solution was drop-casted on graphite (HOPG). In contrast to the sample containing only **OPV** (Fig. 2a), the AFM micrographs of the **pT10–OPV** mixture show uniform small particles with a height of 3–4 nm and a deconvoluted width²⁰ of 5–10 nm (Fig. 2b, d).^{7a} The expected dimensions of the **pT10–OPV** complexes are $\sim 4 \times 4 \times 4$ nm (the length of **pT10** and **OPV** are 3.6 and ~ 4 nm, respectively) and correspond to the size of the objects observed, revealing that the PNA-template controls the size of the **OPV** assemblies.

In conclusion, PNA-templated assemblies have been constructed in MCH of which the size is controlled by the PNA template. This PNA-templated approach can in principle be applied to any functional molecule and makes it possible to construct size-controlled functional nanostructures in organic media. The authors acknowledge K. Pieterse for the artwork and the EURYI scheme for the financial support.

Notes and references

‡ **pT10**:¹³ MALDI-TOF MS (M = 2823.5) m/z = 2824.6 [M + H⁺]. For experimental details and general methods see ref. 7*a*. Sample preparation: **pT10** and **OPV** were dissolved in chloroform. After solvent removal and intensive drying, MCH was added to obtain the appropriate concentration and the solution was heated to 333 K to dissolve both components and slowly cooled. For atomic force microscopy, 2 μ l of the solution at 273 K was drop-cast on freshly cleaved HOPG and allowed to dry in air.

- (a) Special Issue: Tobacco Mosaic Virus: Pioneering Research for a Century, *Philos. Trans. R. Soc. London, Ser. B*, 1999, **354**, 517;
 (b) *Biochemistry*, ed. L. Stryer, 4th Revised edn, W. H. Freeman and Company, New York, United States of America, 1996.
- Examples and reviews of templated polymerizations: (a) J. Gons, E. J. Vorenkamp and G. Challa, J. Polym. Sci., Part A: Polym. Chem., 1975, 13, 1699; (b) T. Inoue and L. E. Orgel, Science, 1983, 219, 859; (c) R. E. Kleiner, Y. Brudno, M. E. Birnbaum and D. R. Liu, J. Am. Chem. Soc., 2008, 130, 4646; (d) R. Saito, Polymer, 2008, 49, 2625; (e) J. C. M. van Hest, Nat. Chem. Biol., 2008, 4, 272.
- Examples of templated self-assembly: (a) J. S. Lindsey, New J. Chem., 1991, 15, 153; (b) T. Sugimoto, T. Suzuki, S. Shinkai and K. Sada, J. Am. Chem. Soc., 2007, 129, 270; (c) S. R. Bull, L. C. Palmer, N. J. Fry, M. A. Greenfield, B. W. Messmore, T. J. Meade and S. I. Stupp, J. Am. Chem. Soc., 2008, 130, 2742; (d) Y. Xu, J. Ye, H. Liu, E. Cheng, Y. Yang, W. Wang, M. Zhao, D. Zhou, D. Liu and R. Fang, Chem. Commun., 2008, 49; (e) A. L. Benvin, Y. Creeger, G. W. Fisher, B. Ballou, A. S. Waggoner and B. A. Armitage, J. Am. Chem. Soc., 2007, 129, 2025.
- 4 F. J. M. Hoeben, P. Jonkheijm, E. W. Meijer and A. P. H. J. Schenning, *Chem. Rev.*, 2005, **105**, 1491.
- 5 Examples of DNA as structural scaffold for particles: (a) Y. Pinto, J. D. Le, N. C. Seeman, K. Musier-Forsyth, T. A. Taton and R. A. Kiehl, *Nano Lett.*, 2005, **5**, 2399; (b) K. V. Gothelf and T. H. LaBean, *Org. Biomol. Chem.*, 2005, **3**, 4023.
- 6 (a) R. Iwaura, F. J. M. Hoeben, M. Masuda, A. P. H. J. Schenning, E. W. Meijer and T. Shimizu, J. Am. Chem. Soc., 2006, 128, 13298; (b) R. Iwaura, K. Yoshida, M. Masuda, M. Ohnishi-Kameyama, M. Yoshida and T. Shimizu, Angew. Chem., Int. Ed., 2003, 42, 1009.
- 7 (a) P. G. A. Janssen, S. Jabbari-Farouji, M. Surin, X. Vila, J. C. Gielen, T. F. A. de Greef, M. R. J. Vos, P. H. H. Bomans, N. A. J. M. Sommerdijk, P. C. M. Christianen, P. Leclère, R. Lazzaroni, P. van der Schoot, E. W. Meijer and A. P. H. J. Schenning, J. Am. Chem. Soc., 2009, 131, 1222; (b) P. G. A. Janssen, J. Vandenbergh, J. L. J. van Dongen, E. W. Meijer and A. P. H. J. Schenning, J. Am. Chem. Soc., 2007, 129, 6078; (c) P. G. A. Janssen, J. L. J. van Dongen, E. W. Meijer and A. P. H. J. Schenning, Chem.-Eur. J., 2009, 15, 352; (d) M. Surin, P. G. A. Janssen, R. Lazzaroni, E. W. Meijer and A. P. H. J. Schenning, Chem.-Eur. J., 2009, 15, 352; (d) M. Surin, P. G. A. Janssen, R. Lazzaroni, E. W. Meijer and A. P. H. J. Schenning, Chem.-Eur. J., 2009, 21, 1126.
- 8 (a) P. K. Lo and H. F. Sleiman, J. Am. Chem. Soc., 2009, 131, 4182; (b) P. K. Lo and H. F. Sleiman, Macromolecules, 2008, 41, 5590.

- 9 Examples of 2D and 3D DNA structures: (a) N. C. Seeman, *Methods Mol. Biol.*, 2005, **303**, 143; (b) F. A. Aldaye, A. L. Palmer and H. F. Sleiman, *Science*, 2008, **321**, 1795.
- 10 It should be noted that DNA has been used as a template in aqueous solvent mixtures, see for example: (a) A. Furstenberg, M. D. Julliard, T. G. Deligeorgiev, N. I. Gadjev, A. A. Vasilev and E. Vauthey, J. Am. Chem. Soc., 2006, 128, 7661; (b) B. A. Armitage, Top. Curr. Chem., 2005, 253, 55.
- (a) P. Jonkheijm, A. Miura, M. Zdanowska, F. J. M. Hoeben, S. de Feyter, A. P. H. J. Schenning, F. C. de Schryver and E. W. Meijer, *Angew. Chem., Int. Ed.*, 2004, 43, 74; (b) F. Würthner, Z. Chen, F. J. M. Hoeben, P. Osswald, C.-C. You, P. Jonkheijm, J. van Herrikhuyzen, A. P. H. J. Schenning, P. P. A. M. van der Schoot, E. W. Meijer, E. H. A. Beckers, S. C. J. Meskers and R. A. J. Janssen, J. Am. Chem. Soc., 2004, 126, 10611.
- 12 (a) P. E. Nielsen and G. Haaima, *Chem. Soc. Rev.*, 1997, 26, 73;
 (b) P. E. Nielsen, *Lett. Pept. Sci.*, 2004, 10, 135;
 (c) M. C. de Koning, G. A. van der Marel and M. Overhand, *Curr. Opin. Chem. Biol.*, 2003, 7, 734.
- 13 E. A. L. Biessen, K. Sliedregt-Bol, P. A. C. T. Hoen, P. Prince, E. van der Bilt, A. R. P. M. Valentijn, N. J. Meeuwenoord, H. Princen, M. K. Bijsterbosch, G. van der Marel, J. H. van Boom and T. J. C. van Berkel, *Bioconjugate Chem.*, 2002, 13, 295.
- 14 MCH was used as apolar solvent to avoid artefacts in the CD measurements, see (a) M. Wolffs, S. J. George, Z. Tomovic, S. C. J. Meskers, A. P. H. J. Schenning and E. W. Meijer, Angew. Chem., Int. Ed., 2007, 46, 8203; (b) S. J. George, Z. Tomovic, M. M. J. Smulders, T. F. A. de Greef, P. E. L. G. Leclere, E. W. Meijer and A. P. H. J. Schenning, Angew. Chem., Int. Ed., 2007, 46, 8206.
- 15 In heptane, the absorption of **OPV** shifts hypsochromically upon self-assembly, see ref. 11*a*.
- 16 It should be noted that λ_{z-c} is not at λ_{max} since the vibronic shoulders also give rise to a Cotton effect.
- 17 (a) P. Jonkheijm, P. van der Schoot, A. P. H. J. Schenning and E. W. Meijer, *Science*, 2006, **313**, 80; (b) M. M. J. Smulders, A. P. H. J. Schenning and E. W. Meijer, *J. Am. Chem. Soc.*, 2008, **130**, 606.
- 18 Remarkably, preliminary molecular modeling simulations reveal disordered pT10–OPV assemblies (see ESI†). Molecular dynamics hint that the assembly of OPV along pT10 is not only driven by H-bonds of the OPV diaminotriazine unit with the thymine bases of pT10 but also with the peptide backbone. Simulations of the CD spectra will be carried out to elucidate the supramolecular organization of the OPVs in these aggregates, see: F. C. Spano, S. C. J. Meskers, E. Hennebicq and D. Beljonne, J. Am. Chem. Soc., 2007, 129, 7044.
- 19 It should be noted that a steeper slope for the non-templated self-assembly curve is obtained near the melting temperature, even though the binding enthalpy is lower. This paradox originates from the fact that although this slope is proportional to the enthalpy change, it has different prefactors for each of the two processes. It is found that the ratio of these slopes curves is approximately given by $\sim \Delta H_e/T_e^{2/}(\eta 1/2)\Delta H_e^{t}/T_e^{12}$ where η is the stoichiometric ratio, equal to one in our experiments. Calculating the ratio of the two slopes according to this equation gives a value of 1.6 which is comparable to the value 1.67 coming directly from the graphs; S. Jabbari-Farouji and P. van der Schoot, manuscript in preparation.
- 20 C. Bustamante, J. Vesenka, C. L. Tang, W. Rees, M. Guthold and R. Keller, *Biochemistry*, 1992, **31**, 22.