



In vitro characterization of an Ab42 specific heptapeptide selected by phage display and preliminary MRI evaluation of Alzheimer's Disease - detection by functionalized USPIO

e-Poster: 545

Congress: ESMRMB 2006 Type: Scientific Poster

Topic: Contrast Agents and Mechanismus

Authors: L. Larbanoix ¹, C. Burtea ¹, S. Laurent ¹, D. Vansthertem ¹, G. Toubeau ¹, F. Van Leuven ², L.

Vander Elst ¹, R.N. Muller ¹; ¹ Mons/BE, ² Leuven/BE

Keywords: MRI, Amyloid-beta, Phage display, Alzheimer

Any information contained in this pdf file is automatically generated from digital material submitted to EPOS by third parties in the form of scientific presentations. References to any names, marks, products, or services of third parties or hypertext links to third-party sites or information are provided solely as a convenience to you and do not in any way constitute or imply ESMRMB's endorsement, sponsorship or recommendation of the third party, information, product, or service. ESMRMB is not responsible for the content of these pages and does not make any representations regarding the content or accuracy of material in this file.

As per copyright regulations, any unauthorised use of the material or parts thereof as well as commercial reproduction or multiple distribution by any traditional or electronically based reproduction/publication method is strictly prohibited.

You agree to defend, indemnify, and hold ESMRMB harmless from and against any and all claims, damages, costs, and expenses, including attorneys' fees, arising from or related to your use of these pages.

Please note: Links to movies, ppt slideshows and any other multimedia files are not available in the pdf version of presentations. http://www.esmrmb.org

1. Purpose

Alzheimer's disease (AD):

- fatal neurodegenerative pathology
- predominant cause of senile dementia
- substantial neuronal loss
- amyloid β(A β) amyloidosis (senile plaques, SP) and tauopathy (neurofibrillary tangles, NFTs).

Although the SP by themselves may not be the direct cause of AD symptoms, a noninvasive method to detect their presence in the brain will allow diagnosis and preventive treatment before occurrence of neurological symptoms and irreversible neurodegeneration.

MRI has a high spatial resolution:

Better suited to assess the SPs in AD, which are often reported to require a spatial resolution of less then 200 micrometers [1]. However, the image acquisition requires high magnetic fields (often higher than 7T) and long acquisition times (several hours) [2-5].

Phage display:

Molecular biology technique that identify high affinity peptides for any molecular target [6].

» This study reports preliminary investigations on a cyclic heptapeptide (P22), which was selected by phage display based on its affinity for A β_{42} [7], and the biochemical and biological characterization of this peptide conjugated to biotin (P22b) or to USPIO (USPIO-P22).

The P22 vectorization of USPIO particles may help to the SP detection by MRI, which has the advantage of a better spatial and anatomical resolution as compared to the imaging techniques of nuclear medicine.

2. Methods and Materials

Conjugation of P22

The disulfide constrained heptapeptide (P22) (NeoMPS, Strasbourg, France) was conjugated either to biotin (P22b) or to USPIO (USPIO-P22) with a linker (8-amino-3,6-dioxaoctanoyl).

Mass spectra were obtained on a Q-tof-2 (Micromass, UK). 0.1 mg of peptide was dissolved in 1 ml of a mixture of formic acid/ acetonitrile/ water (0.5%/ 50%/ 49.5%).

Biotinylated peptide synthesis:

The 8-amino-3,6-dioxaoctanoyl-P22 was purchased from NeoMPS (Strasbourg, France). The peptide was biotinylated as follows: 8 µmol of biotin-NHS (FLUKA, Begium) and 5 µmol of peptide were solubilized in 2 ml of DMF and stirred during 24h at room temperature. 5 ml of water were added, the solution was filtered and dialyzed (cut-off of the membrane: 500; Spectra/Por, VWR, Leuven, Belgium).

The electrospray mass spectrometry confirmed the structure of the biotinylated peptide.

```
ES-MS: [M+H] + : 1439, [M+Na] + : 1461
```

Peptide conjugation to USPIO:

The peptide was linked covalently in 2 steps: the reactive alkyl halogen end of epichlohydrin was first coupled to the hydroxyls of dextran surface of the Fe $_3$ O $_4$ particles to give a terminal glycidyl ether derivative which can be used to link amine containing molecule ([Fig 1] Fig 1).

```
Relaxivity (s ^{-1} mM ^{-1} ): 20 MHz and 37°C: r1 = 32.26 and r2 = 85.32 => r2/r1=2.64 60 MHz and 37°C: r1= 13.50 and r2= 83.75 => r2/r1=6.20
```

In vitro characterization

Evaluation of the dissociation constant (K $_d$) for A β_{42} , as the main component of SP: *ELISA*:

- Immobilization of 100 mg/L of A β_{42} on a polystyrene surface.

- Incubation with a range of concentrations of P22b $[1.25x10^{-6} M 1.91x10^{-9} M]$.
- Detection with peroxydase-conjugated streptavidine (ABC complex) and measurement of OD 405 (nm).

Relaxometry at 60 MHz:

- Incubation of a range of A $~~\beta_{42}$ concentrations [10 $^{-16}$ 10 $^{-7}$ M] with 25 μM of USPIO-P22 in solution.
- Measurement of the transverse relaxation time (T $_2$) at 60 MHz (Bruker mq60 Minispec, Bruker, Karlsruhe, Germany) after 3 minutes of incubation at 37°C.

Blood pharmacokinetics

- Plasma pharmacokinetics were assessed on male Wistar rats anesthetized with 60 mg Nembutal/kg b.w., i.p. The rats were tracheotomized, and the left carotid artery was catheterized for blood collection.
- The contrast agent was injected as a bolus through the femoral vein at a dose of 0.1 mmol Fe/Kg b.w. USPIO has been used as a control.
- Blood samples (~0.3 mL, with saline replacement) were collected before and at 1, 2.5, 5, 15, 30, 45, 60, 90 and 120 min after injection. The iron content of the blood samples was determined by relaxometry at 60 MHz (Bruker mg60 Minispec).
- A two-compartment distribution model was used to calculate the pharmacokinetic parameters such as the elimination half-life (T $_{e1/2}$), the volume of distribution steady state (VD $_{ss}$) and the total clearance (Cl $_{tot}$).

Biodistribution

- The apparent biodistribution has been determined in rats 2h after a single i.v. injection of 0.1 mmol Fe/kg bw.
- The T $_2$ of the organs (liver, kidneys, heart, lungs and brain) was measured by relaxometry at 10 MHz on a Bruker minispec PC-110 (Bruker, Karlsruhe, Germany).
- The R $_2$ values were computed and normalized by the subtraction of R $_2$ values of rat organs that were not exposed to any contrast agent.

In vivo MRI evaluation

Animal model of AD:

- Twenty-two months old double transgenic mice APP[V717I] and PS1-A246E.
- The histology and MRI investigations indicated the presence of amyloid plaques both in the cortex and in the thalamic areas [5] .

Contrast agent administration:

- i.v. injection of 100 micromol Fe/kg of USPIO-P22.
- to open the blood-brain barrier, the mice were first injected with a solution of 25% mannitol i.v. (14mL/kg). The wild type (WT) control mice received the same treatment.

MRI protocol:

- 4.7 T Bruker AVANCE-200 system, vertical bore, micro-imaging device.
- MRI protocol: RARE, TR/TE = 5552/68 ms, NE = 32, matrix 128x128, slice thickness = 1.5 mm, FOV = 2.5 cm, spatial resolution = 195 μ m.

Histology

- After MRI session, brain was dissected, fixed and paraffin-embedded, and slice of 5 µm were cut.
- Incubation with USPIO-P22 (30.4 mM).
- Iron staining with Perl's method (Prussian blue).
- Counter-staining with Sirius Red to localize the SP.

3. Results

In vitro characterization

ELISA:

- P22b has a k $_d$ of 4.13x10 $^{-7}$ M for A β_{42} ([Fig 2] Fig 2).

Relaxometry 60 MHz:

- USPIO-P22 has a K $_{\rm d}$ of 1.2x10 $^{-10}$ M ([Fig 3] Fig 3).

These results shown that the P22 has a very good affinity for its target. The relaxometry seems more sensitive than ELISA, but the difference between the two K $_d$ can also be explained by the different chemical group linked to the peptide or because the A $_{\beta_{42}}$ is in solution when measured by relaxometry and is immobilized in the other case.

Blood pharmacokinetics

- Diminution (p < 0.01) of the blood clearance (Te $_{1/2}$ = 766 min, Cl $_{tot}$ = 0.038 mL/kg/min) of USPIO-P22 as compared to USPIO (Te $_{1/2}$ = 255 min, Cl $_{tot}$ = 0.153 ml/kg/min) ([Fig 4] Fig 4 and [Table 1] Table 1).
- Although not statistically significant, the low VD _{ss} value suggests a diminished diffusion into the interstitial space.

Biodistribution

- The apparent biodistribution of contrast agents was analyzed by measuring T $_2$ values at 10 MHz of the rat organs harvested after pharmacokinetic studies. Because of the intrinsic iron, a normalized R $_2$ (R $_2$ Norm) was computed by subtracting the R $_2$ of organs from rats that did not receive any contrast agent ([Fig 5] Fig 5).
- The contrast agents seem to be localized in liver and lungs, and more USPIO than USPIO-P22 is found in the liver, but the opposite is noticed in the other organs.

Hence, the results suggest a diminished USPIO-P22 uptake by the liver, which could prolong its half-life of elimination.

In vivo MRI evaluation

- The preliminary MRI data on a double transgenic APP/PS1 mouse show a signal decrease in the brain of about 50% after the administration of USPIO-P22 ([Fig 6] Fig 6).
- The effect produced by USPIO-P22 is of only 15% on a control mouse in the same conditions and is comparable to that produced by USPIO.
- The resolution of these images is not adequate to detect individual SP ([Fig 7] Fig 7), but the color map reveals the presence of several dark spots in the thalamic areas and cortex, which could correspond to SP.

Histology

- The SP on histologic sections are stained in red with Sirius Red ([Fig 8] Fig 8).
- On the slices incubated with USPIO-P22, a blue iron staining is visible around the cores of SP.
- The negative control with USPIO does not show any blue iron staining around the SP.

Therefore, USPIO-P22 seems to interact specifically with amyloid fibers. The core of SP is too dense to allow the penetration of the contrast agent deep inside.

4. Conclusion

The contrast agent USPIO-P22 has a good affinity for A β_{42} and seems to be specific to SP. It could represent a potential tool for AD diagnosis after further characterization.

5. References

- 1. Poduslo J.F., Curran G., Peterson J., McCormick D., Fauq A., Khan M., and Wengenack T. Design and chemical synthesis of a magnetic resonance contrast agent for imaging Alzheimer's disease amyloid plaques. Neurobiol Aging 2004;25:S56-S57.
- 2. Jack, C.R.; Wengenack, T.M.; Reyes, D.A.; Garwood, M.; Curran, G.L.; Borowski, B.J.; Lin, J.; Preboske, G.M.; Holasek, S.S.; Adriany, G.; Poduslo, J.F. In vivo magnetic resonance microimaging of individual amyloid plaques in Alzheimer's transgenic mice. J Neurosci. 2005;25:10041-10048.
- 3. Benveniste H., Einstein G., Kim K.R., Hulette C., and Johnson A. Detection of neuritic plaques in Alzheimer's disease by magnetic resonance microscopy . Proc Natl Acad Sci U S A. 1999;96:14079-14084.
- 4. Zhang J., Yarowsky P., Gordon M.N., Di Carlo G., Munireddy S., van Zijl P.C.M., and Mori S. Detection of amyloid plaques in mouse models of Alzheimer's disease by magnetic resonance imaging. Magn Reson Med. 2004;51:452-457.
- 5. Vanhoutte G., Dewachter I., Borghgraef P., Van Leuven F., and Van der Linden A. Noninvasive in vivo MRI detection of neuritic plaques associated with iron in APP[V717I] transgenic mice, a model for Alzheimer's disease. Magn Reson Med. 2005;53:607-613.

- 6. Smith G.P. and Petrenko V.A. Phage display. Chem Rev. 1997;97:391-410.
- 7. Larbanoix L, Burtea C, Laurent S, Vander Elst L, Muller RN, ESMRMB, 2005, Basel, Swiss.

6. Personal Information

Lionel Larbanoix
Department of Organic and Biomedical Chemistry
NMR and Molecular Imaging Laboratory
University of Mons-Hainaut, B-7000 Mons, Belgium
http://w3.umh.ac.be/ ~nmrlab/

tel: +32-65-373519 fax: +32-65-373520

7. Mediafiles:

Fig 1

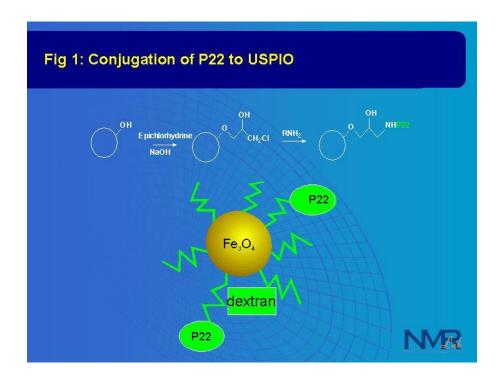


Fig 2

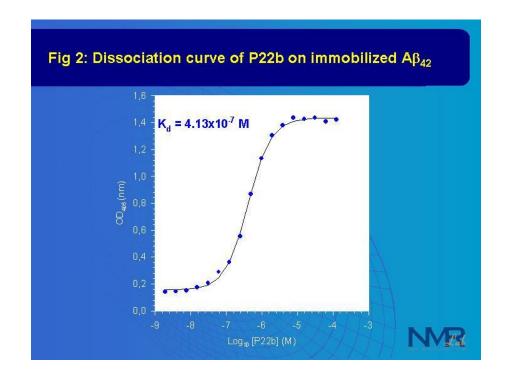


Fig 3

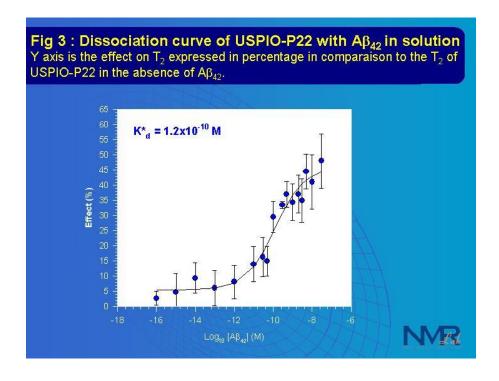


Fig 4

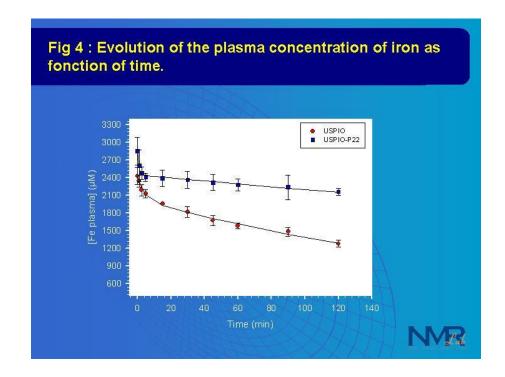


Fig 5

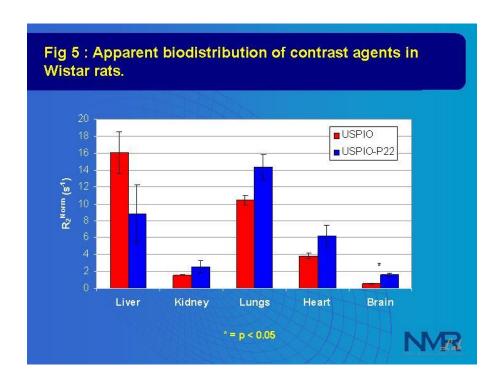


Fig 6

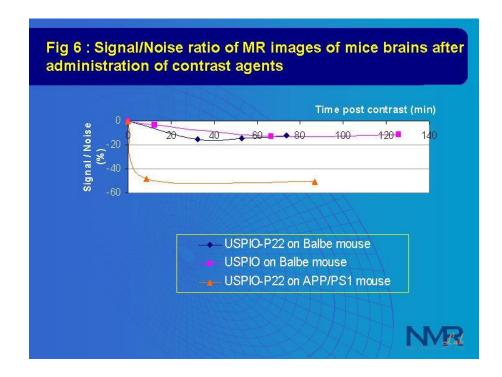


Fig 7

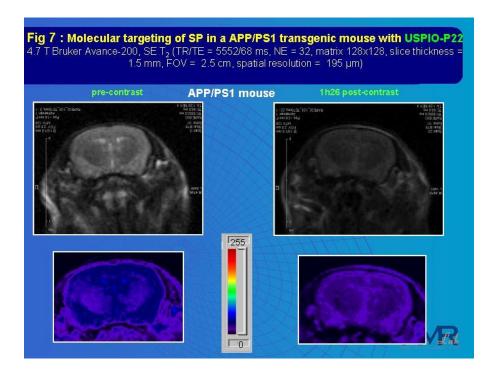


Fig 8

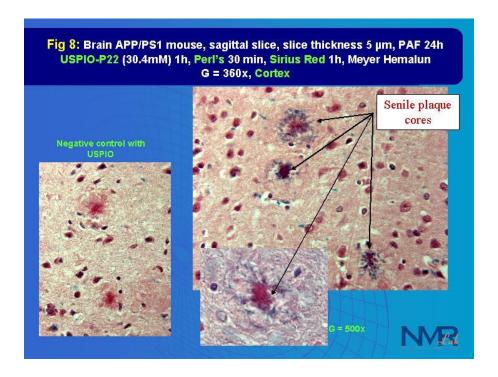


Table 1

