

A promising phospholipase A2-targeted peptide slowing amyloid beta pathology in an Alzheimer's disease mouse model

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health



INTRODUCTION

Negative control

Alzheimer disease (AD)

- Most common dementia worldwide with 2 main features : extracellular senile plaques of amyloid β (A β) & intracellular neurofibrillary tangles of tau protein
- Since 1993, no new drug was approved by FDA (> 95% failed during clinical trials [1])
- Actual therapies are only symptomatic & do not slow the progression of the disease

AD management

- Only symptomatic treatments allowing a better quality of life for patients
- Various phospholipase isoforms involved in memory impairment and neurodegeneration in AD
- PLA₂ signaling pathway involved in AD [2] \rightarrow PLA₂ inhibition has shown neuron protection against apoptosis induced by A β [3]



Figure 1. Our hypothesis concerning the progress of the AD process: accumulation of Aβ at the cerebral level results in excessive stimulation of NMDAR, triggering the exacerbated influx of Ca²⁺ which activates kinases responsible of PLA₂ activation. Arachidonic acid (AA) produced promotes synaptic function disruption, tau phosphorylation and aggregation, increased A^β production and ROS generation, as well as neuro-inflammation and apoptotic death of neurons

METHODS

- Inhibitory potential of PL-P25: Pre-incubation of differentiated NT2/D1 cells during 30 minutes with peptides (20 μ M) before induction with glutamate (50 μ M) \rightarrow AA dosage (AA ELISA kit, Cusabio, USA).
- **Immunofluorescence:** subcellular localization of PLA₂ and p-PLA₂, of AA-depending enzymes such as COX2, ALOX5, and beta-actin.
- In vivo molecular imaging: APP/PS1 mice (Jackson Laboratory, Maine, USA) were injected with 200µmol Fe/kg b.w of USPIO-PHO [4]. Then, images were acquired at the level of the head with T_2 -weighted RARE imaging protocol (TR/TE = 3000/60 ms, RARE factor = 4, NEX = 6, matrix = 512x512, FOV = 2.5cm, slice thickness 1mm, 20 axial slices, spatial resolution = 48µm, TA = 38m24sec)
- Barnes maze: study of the spatial memory of non treated healthy mice and APP/PS1 mice during the period of treatment with PLP25-LRP2 or NSP (1.5 month). All performances were recorded and analyzed manually.

ALOX5 in NT2/D1

Immunohistochemistry: detection of AP, APP, PLA₂, NMDAR and p-tau

Development of a therapeutic strategy by targeting a key actor in the phospholipase (PLA₂) signalling involved in AD using a peptide identified by phage display (PLP25) and rendered able to cross the BBB by coupling to a vector peptide (LRP2) targeting the LDLR.



Figure 2. The glutamate shows a stimulatory effect on PLA₂ (positive control) compared to the negative control (non-induced and non-inhibited cells), attested by a significant increase in the released AA. PLP25-LRP2 complex significantly decreases the released AA levels, whereas the non-specific peptide (NSP) shows no effect (*: p<0.05; **: p<0.01).



Figure 5. Study of the spatial memory of APP/PS1 mice by the Barnes maze. Mice treated with PLP25-LRP2 generally made fewer errors (A) and spent more time in the correct quadrant (B) than the non-treated or NSP-treated mice.



Positive control

Figure 3. Detection by IF of (A) glutamate-induced PLA₂ and (B) Aβ-induced PLA₂ on neuron differentiated NT2/D1 cells (NT2/D1_n) highlighted by Dylight 594 (red). Nuclei appear blue using DAPI. PLA₂ migrates to neurites in both stimulations. In both cases, PLP25-LRP2

Negative contrast

experiments



Figure 4. Study of the inhibitory effect of PLP25 on cellular biomarkers linked to PLA₂. PLA₂ stimulation by glutamate results in its phosphorylation and migration to membranes and neurites, the activation of COX2 and/or ALOX5 depending on the cell type, the translocation of these enzymes into cell processes and the restructuration of the beta-actin cytoskeleton. PLP25 prevents all these phenomena by its interaction with PLA_2 . * p < 0.05



Phosphorylated PLA₂ in NT2/D1



Figure 7. (A) Detection of amyloid plaques (AP) by IHC on brain slices of healthy NMRI and APP/PS1 mice, highlighted by brown spots. Figure 8. Detection of amyloid precursor protein (APP) by IHC. The After 1.5 months of treatment, the injection of PLP₂₅-LRP₂ to APP/PS1 mice reduced the number and size of AP in contrast to NSP labeling obtained for PLP25-LRP2-treated mice is intermediate between injection, whereas older and non-treated mice show AP progression. (B) AP were counted manually in the whole brain and the the 10 months-diseased mice (more cytoplasmic) and healthy mice hippocampus of mice treated with PLP₂₅-LRP₂ or NSP, and in older APP/PS1. (mainly membranous) with a closer approach to healthy mice.

RESULTS





NT2/D1_n







Figure 9. Detection of PLA₂ by IHC. Pathological conditions induce the overexpression of PLA₂ in both the cortex and the hippocampus. Mice treated with PLP25-LRP2 have a clearly weak overall labeling, unlike mice treated with the NSP, in the range of diseased mice.

Figure 10. Similar detection of NMDAR by IHC reveals a decreased expression of NMDAR in diseased mice. The treatment with PLP25-LRP2 allows to increase its expression, unlike the treatment with NSP.

Figure 11. Detection of phospho-tau (p-Tau) by IHC. Healthy mice and mice treated with PLP25-LRP2 have shown a p-Tau labeling mainly in the cell bodies and in the nucleus, whereas in diseased mice and NSPtreated mice, p-Tau is located in axons.



Prevent COX2 and ALOX5 translocation after Glu stimulation

EX VIVO

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> Rotary Espoir en tête



IHC on mice brain slices showed that PLP₂₅-LRP₂ injection allows to restore in the range of healthy mice the expression, cellular localization and activity of selected biomarkers of interest for AD.

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